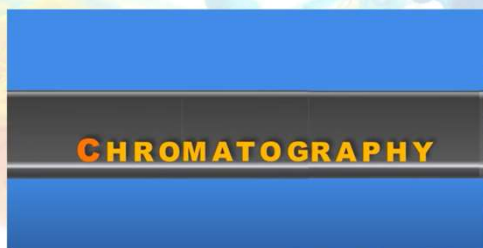


# Chromatography

Components partition between two phases.

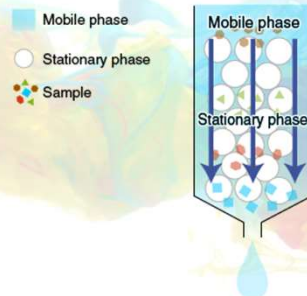


1

<https://www.youtube.com/watch?v=0m8bWK4mRMM>

## Chromatography

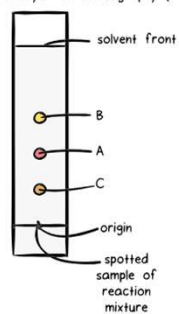
The principle and method



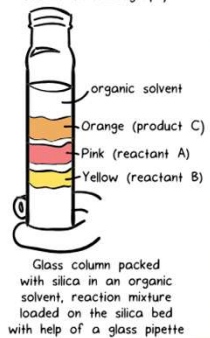
2

<http://uo.mbl.co.jp/biole/support/method/chromatography.html>

Thin Layer Chromatography (TLC)

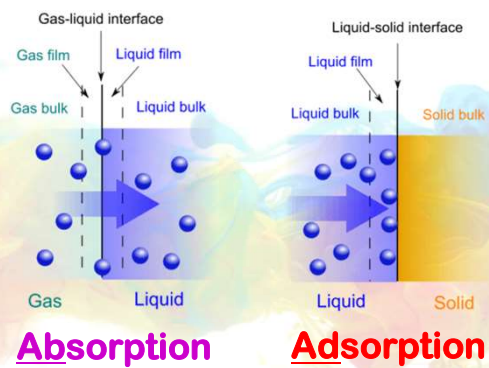


Column Chromatography



3

<https://www.khanacademy.org/science/class-11-chemistry-india/a/xb66cb8fc2bd00c8:in-in-organic-chemistry-some-basic-principles-and-techniques/xb66cb8fc2bd00c8:in-in-methods-of-purification-of-organic-compounds/a/principles-of-chromatography>



**Absorption**

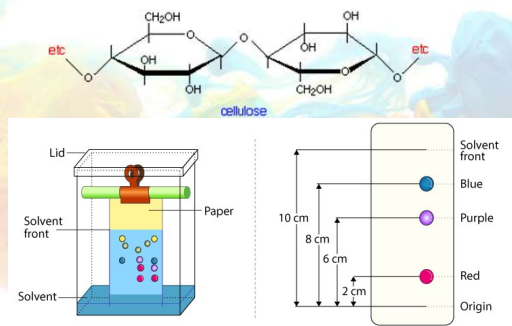
**Adsorption**

[https://www.diffen.com/difference/Absorption\\_vs\\_Adsorption](https://www.diffen.com/difference/Absorption_vs_Adsorption)

4

4

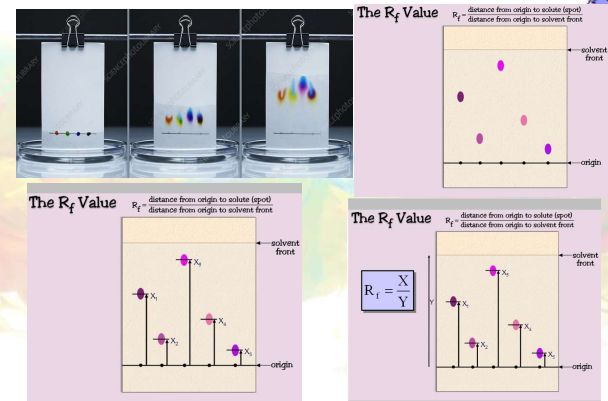
Paper is made of cellulose fibres, and cellulose is a polymer of the simple sugar, glucose.



<https://byjus.com/chemistry/paper-chromatography/>

5

## Paper chromatography



<https://www.sciencemag.com/media/860110/view/paper-chromatography>

<https://slideplayer.com/slide/4701853/>

6

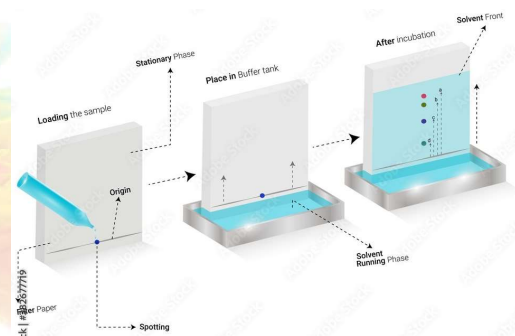
Table 12.5.1 . Properties of HPLC Mobile Phases

| mobile phase           | polarity index ( $P'$ ) | UV cutoff (nm) |
|------------------------|-------------------------|----------------|
| cyclohexane            | 0.04                    | 210            |
| <i>n</i> -hexane       | 0.1                     | 210            |
| carbon tetrachloride   | 1.6                     | 265            |
| <i>i</i> -propyl ether | 2.4                     | 220            |
| toluene                | 2.4                     | 286            |
| diethyl ether          | 2.8                     | 218            |
| tetrahydrofuran        | 4.0                     | 220            |
| ethanol                | 4.3                     | 210            |
| ethyl acetate          | 4.4                     | 255            |
| dioxane                | 4.8                     | 215            |
| methanol               | 5.1                     | 210            |
| acetonitrile           | 5.8                     | 190            |
| water                  | 10.2                    | —              |

[https://chem.libretexts.org/Bookshelves/Analytical\\_Chemistry/Analytical\\_Chemistry\\_2.1\\_\(Harvey\)/12%3A\\_Chromatographic\\_and\\_Electrophoretic\\_Methods/12.05%3A\\_High-Performance\\_Liquid\\_Chromatography](https://chem.libretexts.org/Bookshelves/Analytical_Chemistry/Analytical_Chemistry_2.1_(Harvey)/12%3A_Chromatographic_and_Electrophoretic_Methods/12.05%3A_High-Performance_Liquid_Chromatography)

7

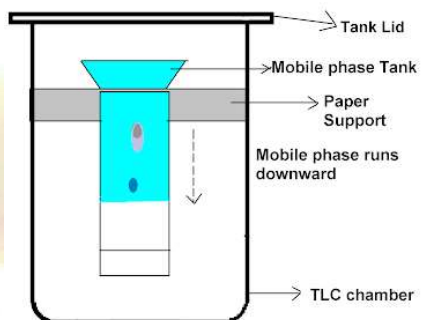
## Ascending techniques



<https://stock.adobe.com/it/images/ascending-chromatography-schematic-diagram-analysis-of-amino-acids-paper-chromatography-technique-biochemical-techniques/382677719>

8

### Descending techniques

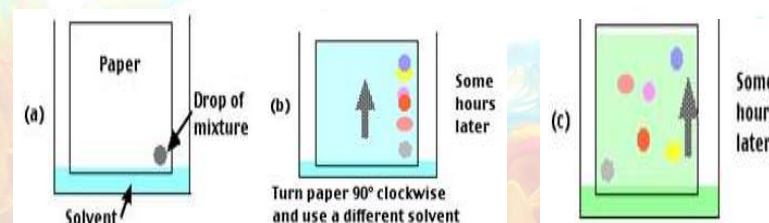


<https://pharmainformationzone.blogspot.com/2020/04/descending-chromatography.html>

9

9

### Two-dimensional chromatography (2D)



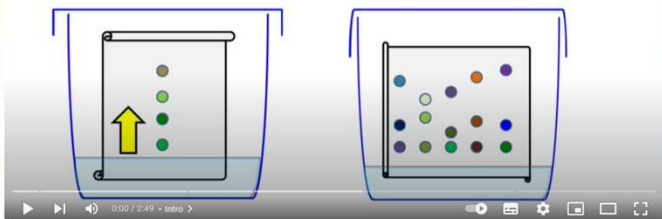
<http://chromatographyscience.blogspot.com/2012/08/two-dimensional-techniques-in-paper.html>

10

10

**Quickly  
understand**

### Paper Chromatography

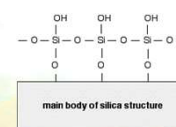


[https://www.youtube.com/watch?v=mz\\_xcN7TK\\_U](https://www.youtube.com/watch?v=mz_xcN7TK_U)

11

11

### Thin layer chromatography



<https://mz-at.de/en/chromatography/further-categories/thin-layer-chromatography/>  
<http://www.chemguide.co.uk/analysis/chromatography/thinlayer.html#top>

<https://www.linkedin.com/pulse/thin-layer-chromatography-tlc-market-size-evaluating>

12

12

The distance from the origin to the compound.  
 $R_f = \frac{\text{The distance from the origin to the solvent front.}}{\text{The distance from the origin to the solvent front.}}$

Aspirin  $R_f = 0.5$   
 2-Hydroxybenzoic acid  $R_f = 0.75$ .

Key  
 A: Starting material  
 B: Crude product  
 C: Recrystallised product

Using fluorescence

<https://www.careerpower.in/school/chemistry/thin-layer-chromatography>

<https://www.chegg.com/homework-help/questions-and-answers/see-two-spots-crude-product-thin-layer-chromatography-aspirin-q62594548>

13

### Comparison of different extraction methods: solvent extraction and steam distillation used for the analysis of the volatile components in *M. cajuputi* leaves

Mr. Suphachai Nampho

สารสกัดที่ได้จากการสกัดด้วยตัวทำละลาย

น้ำมันหอมระเหยจากการกลั่นด้วยไอน้ำ

14

ผลการแยกสารสกัดตัวอย่างในอัตราส่วนเฟสเคลื่อนที่ต่าง ๆ (สเกลเซนต์มทานอล)

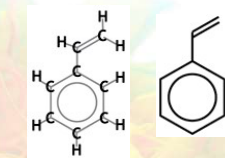
15

### Separation of Plant Pigments by Column Chromatography (CC)

<https://byjus.com/chemistry/column-chromatography/>

16

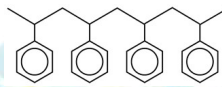
## Ion Exchange Chromatography



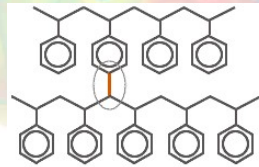
Chemical formula of styrene



Divinylbenzene (DVB)



A small fraction of a polystyrene chain



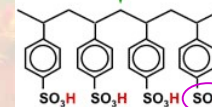
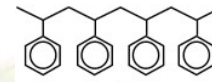
Cross-linked polystyrene

[http://dardel.info/IX/resin\\_structure.html](http://dardel.info/IX/resin_structure.html)

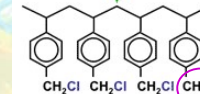
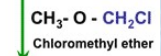
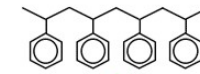
17

17

## Resin Structure and Manufacture



- **Cation Exchange**
  - Sulfonic acid
  - $\text{SO}_3^- (\text{H}^+)$
  - Carboxylic Acid

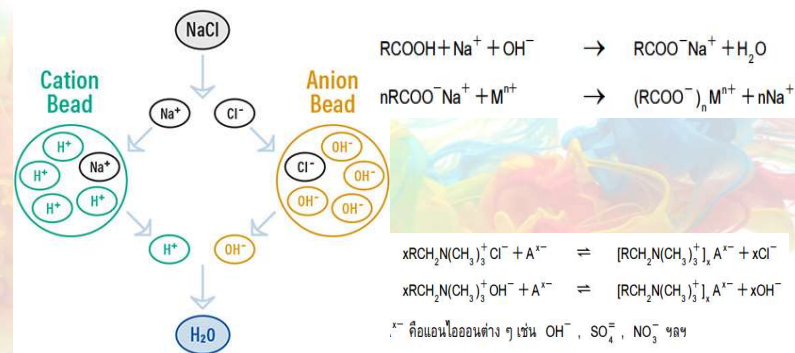


- **Anion Exchange**
  - Primary Amine
  - $\text{NH}_3^+(\text{OH}^-)$
  - Tertiary Amine

[http://dardel.info/IX/resin\\_structure.html](http://dardel.info/IX/resin_structure.html)

18

## Resin Structure and Manufacture



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## Mobile Phase

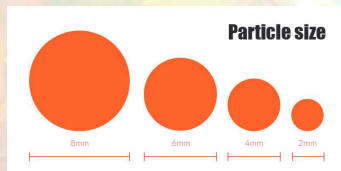
- Cation Exchange -  $\text{H}^+$
- Anion Exchange -  $\text{OH}^-$
- pH is controlled with a Buffer Solution
  - e.g.  $\text{HCO}_3^-/\text{CO}_3^{2-}$

20

20

## ปัจจัยที่ส่งผลต่อเรซิน

## 1. size of particles



## 2. Degree of cross-linking

| % Crosslinking | Moisture (%) | Capacity (meq/ml) |
|----------------|--------------|-------------------|
| 2              | 71           | 1.2               |
| 4              | 61           | 1.5               |
| 6              | 53           | 1.8               |
| 8              | 46           | 2.0               |
| 10             | 42           | 2.2               |
| 12             | 37           | 2.4               |
| 14             | 34           | 2.6               |
| 16             | 32           | 2.8               |

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## ปัจจัยที่ส่งผลต่อเรซิน

## 3. Strength of functional group

## 4. Number of functional group

| Resin | Functional group                                                                                                                      | Weak or Strong | Functional pH Range | Nature          | Functional group                                                                | Range  | Applications          | Capacity meq/g | Commercial resins              |
|-------|---------------------------------------------------------------------------------------------------------------------------------------|----------------|---------------------|-----------------|---------------------------------------------------------------------------------|--------|-----------------------|----------------|--------------------------------|
| DEAE  | diethylaminoethyl<br>[-N <sup>+</sup> (C <sub>2</sub> H <sub>5</sub> ) <sub>2</sub> H <sup>+</sup> ]                                  | weak anion     | pH 2 - 9            | Strong acid     | -SO <sub>3</sub> H                                                              | pH < 2 | Water softening       | 5              | Lewatit S100, Amberlite IR-200 |
| ANX   | diethylaminopropyl<br>[-N <sup>+</sup> (C <sub>2</sub> H <sub>5</sub> ) <sub>2</sub> H <sup>+</sup> ]                                 | weak anion     | pH 2 - 9            | Weak acid       | =PO <sub>3</sub> H                                                              | pH > 4 | Rare earth processing | 10             | Amberlite IFC-84 Duolite CC3   |
| Q     | quaternary amine<br>[-N <sup>+</sup> (CH <sub>3</sub> ) <sub>4</sub> ]                                                                | strong anion   | pH 1 - 14           |                 |                                                                                 |        |                       |                |                                |
| CM    | carboxymethyl<br>[-O-CH <sub>2</sub> COO <sup>-</sup> ]                                                                               | weak cation    | pH 5 - 10           | Strong base     | -NR <sub>3</sub> X                                                              | 0 - 14 | Uranium processing    | 4              | Zerolit MPF Lewatit MP200      |
| S     | methyl sulfonate<br>[O-CH <sub>2</sub> -CHOH-CH <sub>2</sub> -O-CH <sub>2</sub> -CHOH-CH <sub>2</sub> -SO <sub>3</sub> <sup>-</sup> ] | strong cation  | pH 2 - 12           | Weak base       | -NH <sub>2</sub> , -NR <sub>2</sub> , -NHR                                      | 0 - 10 | Uranium processing    | 5              | Zerolit MPH Lewatit MP62       |
| SP    | sulphonyl group<br>[-CH <sub>2</sub> -CH <sub>2</sub> -CH <sub>2</sub> SO <sub>3</sub> <sup>-</sup> ]                                 | strong cation  | pH 2 - 14           | Chelating resin | Picolylamine<br>-N(CH <sub>2</sub> C <sub>6</sub> H <sub>4</sub> ) <sub>2</sub> | -      | Cu & Ni removal       | 1.5            | XFS 4195                       |

22

22

[https://www.researchgate.net/figure/1-Classification-of-ion-exchange-resins-based-on-functional-groups\\_tbl1\\_232957574](https://www.researchgate.net/figure/1-Classification-of-ion-exchange-resins-based-on-functional-groups_tbl1_232957574)

## Ion Exchange Chromatography – General Characteristics

- In general, ion exchangers favor binding of ions with:

- ↑ charge
  - 3<sup>+</sup> > 2<sup>+</sup> > 1<sup>+</sup> ; Na<sup>+</sup> < Ca<sup>2+</sup> < La<sup>3+</sup> < Th<sup>4+</sup>
- ↓ hydrated radius
  - H<sup>+</sup> > Li<sup>+</sup> > Na<sup>+</sup> > K<sup>+</sup>
- ↓ hydrated; exchange ↑
- ↑ polarizability

| Ion              | Ionic Hydration Radius/Å |
|------------------|--------------------------|
| Li <sup>+</sup>  | 3.82                     |
| Na <sup>+</sup>  | 3.58                     |
| K <sup>+</sup>   | 3.31                     |
| Rb <sup>+</sup>  | 3.29                     |
| Cs <sup>+</sup>  | 3.29                     |
| Ca <sup>2+</sup> | 4.12                     |
| Mg <sup>2+</sup> | 4.28                     |
| Sr <sup>2+</sup> | 4.12                     |

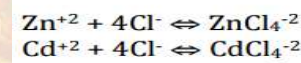
[https://www.researchgate.net/figure/The-hydration-radius-and-potential-of-the-ions\\_tbl1\\_370169111](https://www.researchgate.net/figure/The-hydration-radius-and-potential-of-the-ions_tbl1_370169111)

23

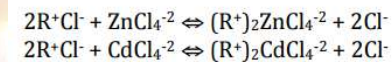
23

## Separation of Cadmium and Zinc on an Anion-Exchange Resin, and Their Determination Using Standardized EDTA Solution

| Metal ions       | Ionic radius (Å) | Hydrated radius (Å) |
|------------------|------------------|---------------------|
| Zn <sup>2+</sup> | 0.83             | 4.30                |
| Cd <sup>2+</sup> | 1.03             | 4.26                |



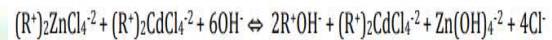
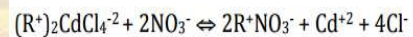
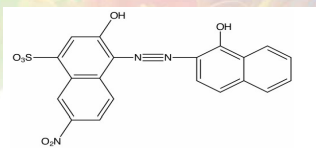
## Sample Addition to Anion-Exchange Resin:



[https://www.researchgate.net/figure/Radius-and-hydration-energy-for-Zn-2-and-Cd-2-18\\_tbl1\\_232987565](https://www.researchgate.net/figure/Radius-and-hydration-energy-for-Zn-2-and-Cd-2-18_tbl1_232987565)

24

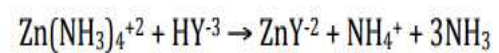
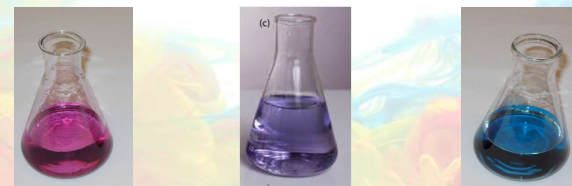
24

**Elution of Zinc:****Elution of cadmium:****Eriochrome black T (EBT) indicator**

25

25

The titration reaction between the zinc tetra amine complex and  $HY^{-3}$  (the form of EDTA predominating at pH10)



26

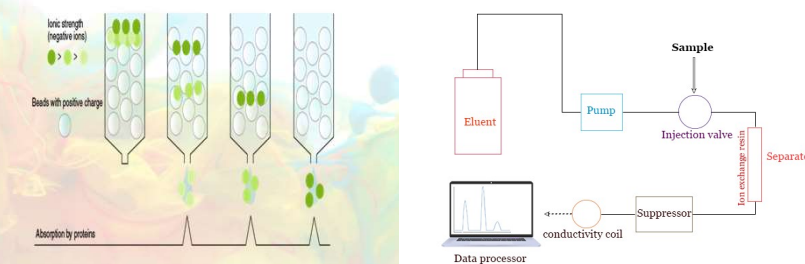
26

**Ion Exchange Chromatography**

<https://www.youtube.com/watch?v=H4U4ndf2ayg>

27

27

**Ion-exchange chromatography****Instrumentation of Ion exchange**

<http://suu.mbl.co.jp/bio/e/support/method/chromatography.html>

<https://www.priyamstudycentre.com/2021/10/ion-exchange-chromatography.html>

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28

## Gas Chromatography



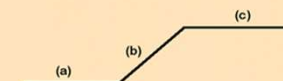
<https://www.youtube.com/watch?v=IX25exzwKhl>

29

29

## Isothermal & Temperature programming

### A temperature program

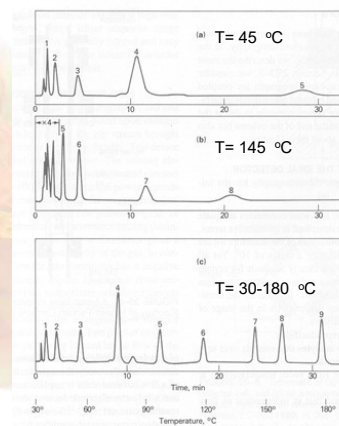


a - initial temperature and time

b - ramp (°C/min)

c - final hold time and temperature

Some GCs will allow for a more complex program.



<https://slideplayer.com/slide/9198088/>

30

30

## High performance liquid chromatography

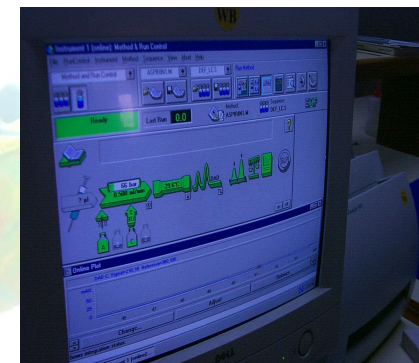


<https://www.youtube.com/watch?v=eGjocRUvJg>

31

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## HPLC



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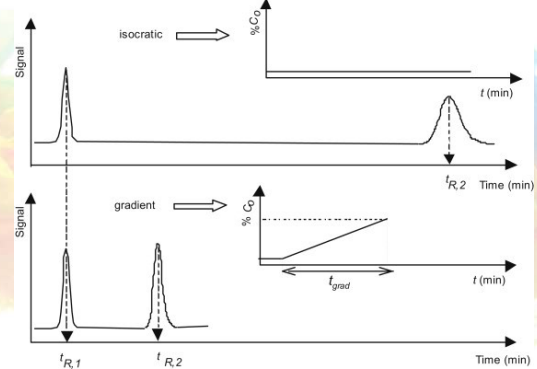
32

## High performance liquid chromatography

|                     | Normal Phase                         | Reversed Phase                |
|---------------------|--------------------------------------|-------------------------------|
| Stationary phase    | Polar (silica gel)                   | Non-polar (C18)               |
| Mobile phase        | Non-polar (organic solvents)         | Polar (aqueous/organic)       |
| Sample movement     | Non-polar fastest                    | Polar fastest                 |
| Separation based on | Different polarities (functionality) | Different hydrocarbon content |

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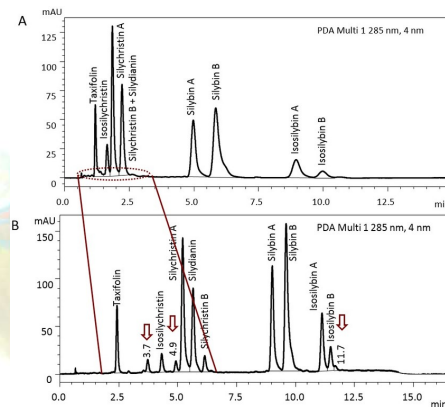
## Isocratic & Gradient elution



<https://www.sciencedirect.com/topics/materials-science/gradient-elution>

34

34

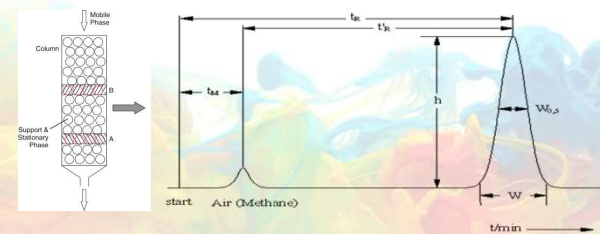


Chromolith RP-18e (100 × 3 mm) column; isocratic conditions: Mobile phase: 2% acetonitrile, 37% methanol, 0.1% formic acid, flow rate 1.1 mL/min,  $t = 25^\circ\text{C}$ ; gradient conditions: Mobile phase: A = 5% acetonitrile, 0.1% formic acid; B = 80% methanol, 0.1% formic acid; gradient: 0 min 30% B, 12 min 60% B, 13 min 60% B, 14 min 30% B, 16.5 min stop, flow rate 1.1 mL/min,  $t = 25^\circ\text{C}$ .

[https://www.researchgate.net/figure/Comparison-of-the-isocratic-A-and-gradient-B-HPLC-analytical-method-for-silymarin-SM\\_fig2\\_338806376](https://www.researchgate.net/figure/Comparison-of-the-isocratic-A-and-gradient-B-HPLC-analytical-method-for-silymarin-SM_fig2_338806376)

35

## The Chromatogram



$t_M$ : retention time of air or mobile phase

$t_R$ : retention time

<https://slideplayer.com/slide/9221930/>

<https://www.sciencedirect.com/topics/chemistry/chromatography>

36

36

33

### The Capacity Factor; $k'$



$$k'_A = \frac{K_A V_S}{V_M} \dots\dots\dots(1)$$

$$k'_A = \frac{t_R - t_M}{t_M} \dots\dots\dots(2)$$

$k'_A$  : capacity factor

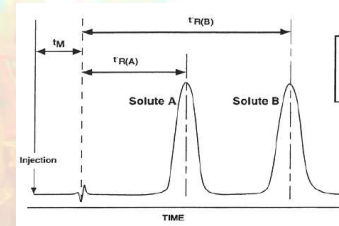
$V_S$  : volume of stationary phase

$K_A$  : partition ratio

$V_M$  : volume of mobile phase

37

### The Selective Factor; $\alpha$

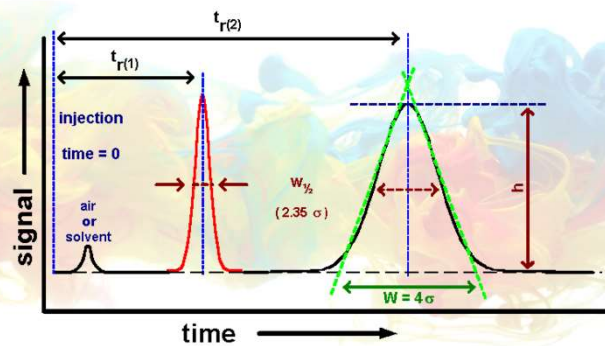


$$\alpha = \frac{K_B}{K_A} \dots\dots\dots(3)$$

$$\alpha = \frac{k'_B}{k'_A} \dots\dots\dots(4)$$

$$\alpha = \frac{(t_R)_B - t_M}{(t_R)_A - t_M} \dots\dots\dots(5)$$

38



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### The Efficiency of chromatographic columns

*Number of theoretical plates;  $N$*

$$N = \frac{L}{H} \dots\dots\dots(6)$$

$$N = 16 \left( \frac{t_R}{w} \right)^2 = 5.54 \left( \frac{t_R}{w_{1/2}} \right)^2 \dots\dots\dots(7)$$

$L$  : the length of the column

$H$  : plate height

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**Number of theoretical plates;  $N$**

$$N_{req} = 16R^2 \left( \frac{\alpha}{\alpha - 1} \right)^2 \left( \frac{1 + k'_2}{k'_2} \right)^2$$

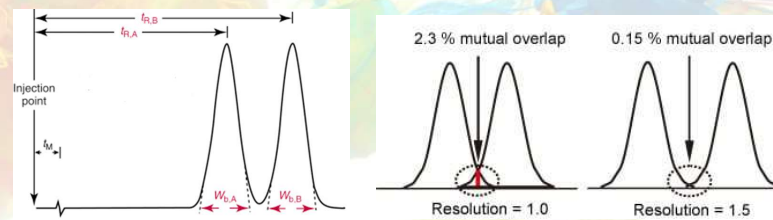
$$k'_2 = \frac{t_{R2} - t_M}{t_M}$$

41

**Resolution;  $R$**



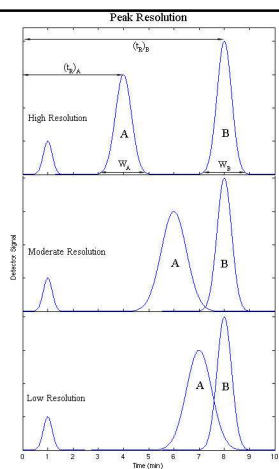
$$R = \frac{t_{R2} - t_{R1}}{\frac{w_1}{2} + \frac{w_2}{2}} = \frac{2(t_{R2} - t_{R1})}{w_1 + w_2} \dots\dots\dots(8)$$



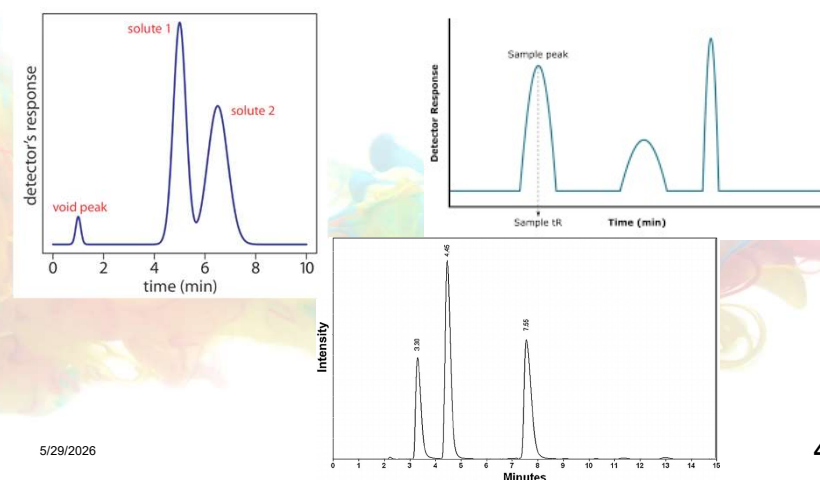
<https://www.sciencedirect.com/topics/chemistry/chromatography>

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**Resolution**



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### ตัวอย่างที่ 9.1

การวิเคราะห์วิตามิน B6 ในน้ำผึ้งด้วย HPLC โดยใช้สารมาตรฐานที่มีความเข้มข้น 5.00 mg/L ได้พื้นที่พีค (area) = 1200 หน่วย เมื่อนำตัวอย่างไม่ทราบความเข้มข้น ได้พื้นที่พีค = 1560 หน่วย ความเข้มข้นของวิตามิน B6 ในน้ำผึ้งเท่ากับกี่ mg/L (สมมติความสัมพันธ์เป็นเชิงเส้นตรง)

### ตัวอย่างที่ 9.2

ในการวิเคราะห์สารลูทีนในเมล็ดข้าวด้วย HPLC:

- การสกัดสาร 1 กรัมด้วยเมทานอล 3.00 มล.
- สร้างกราฟมาตรฐานได้สมการเชิงเส้นคือ  $y = 37.857x + 14.838$
- กรองสารสกัดได้ 1.50 มล. นำไปวิเคราะห์ ได้พื้นที่ใต้พีคเท่ากับ 2541 สารลูทีนในเมล็ดข้าวก็มีผลกี่กรัม

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### ตัวอย่างที่ 9.3

จากการวิเคราะห์กรดแกลลิกในพืชสมุนไพรพื้นบ้าน ชิง ข่า ตะไคร้ เมื่อนำมาสร้างกราฟมาตรฐานโดยพล็อตระหว่างความเข้มของสารมาตรฐานแกลลิกกับพื้นที่ใต้พีคได้ผลดังตาราง

จงสร้างกราฟมาตรฐาน และเมื่อนำตัวอย่างมาวิเคราะห์ได้พื้นที่ใต้พีคดังนี้

1. ชิง เท่ากับ 524
2. ข่า เท่ากับ 890
3. ตะไคร้ เท่ากับ 450

โดยสกัดสาร 1 กรัมในเอทานอล 2.00 มล. ในตัวอย่างมีกรดแกลลิก (MW = 170.12 g/mol) กี่มิลลิกรัมในตัวอย่าง 1 กิโลกรัม

| ความเข้มข้น | พื้นที่ใต้พีค |
|-------------|---------------|
| 0.5         | 28.27         |
| 1           | 47.80         |
| 5           | 158.53        |
| 10          | 370.70        |
| 30          | 1152.48       |
| 60          | 2250.80       |
| 100         | 4044.95       |
| 200         | 7480.62       |

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### ตัวอย่างที่ 9.4

จากโครมาโทแกรมของ HPLC แสดงพีคของสาร 2 ตัวที่ตรวจพบดังนี้:

1. ค่า  $k'$  ของสาร A และ B
2. ค่า **selectivity** ( $\alpha$ ) ของ B เทียบกับ A
3. ค่า **resolution** ระหว่าง A กับ B

| รายการ                       | สาร A    | สาร B    |
|------------------------------|----------|----------|
| Retention time ( $t_R$ )     | 2.31 min | 5.64 min |
| ความกว้างพีคที่ฐาน ( $w_b$ ) | 0.50 min | 0.60 min |
| เวลา dead time ( $t_0$ )     | 1.0 min  | 1.0 min  |

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### ตัวอย่างที่ 9.5

จากโครมาโทแกรมของการวิเคราะห์ด้วย HPLC พบว่า:

- สาร C มี  $t_R = 8.20$  min
- ความกว้างของพีคที่ครึ่งความสูง ( $w_{1/2}$ ) = 0.40 min
- ความยาวคอลัมน์ = 300 เมตร

1. คำนวณ จำนวน theoretical plates (N) โดยใช้สูตรจาก  $w_{1/2}$
2. คำนวณ plate height (H) ของคอลัมน์นี้

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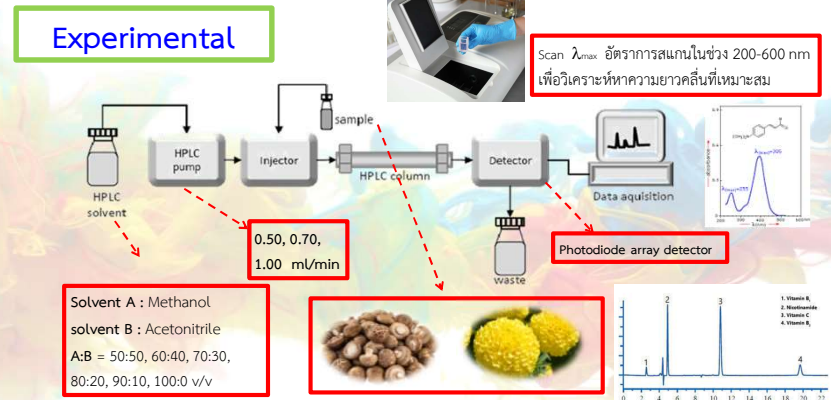
## DEVELOPMENT OF A HIGH-PERFORMANCE LIQUID CHROMATOGRAPHY METHOD FOR THE QUANTITATIVE DETERMINATION OF RUTIN AND LUTEIN IN FLOWERS AND MUSHROOMS

นางสาวเกศนีนีภา โชวแข็ง  
รหัสนักศึกษา 6504103304 สาขาวิชาเคมี  
อาจารย์ที่ปรึกษา ผศ.ดร.ศิริรัตน์ โพศาลสุทธิชล

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### Experimental



Scan  $\lambda_{max}$  อัตราการสแกนในช่วง 200-600 nm เพื่อวิเคราะห์หาความยาวคลื่นที่เหมาะสม

0.50, 0.70, 1.00 mL/min

Solvent A : Methanol  
solvent B : Acetonitrile  
A:B = 50:50, 60:40, 70:30, 80:20, 90:10, 100:0 v/v

Photodiode array detector

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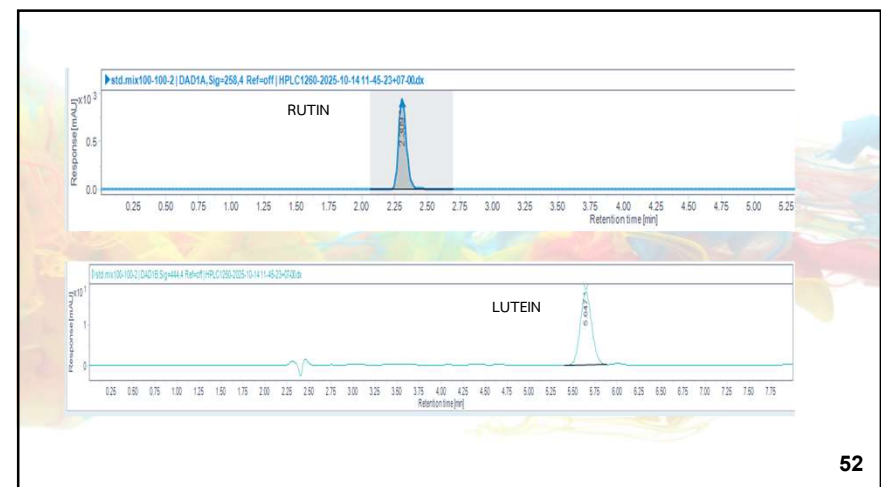
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### The optimize Condition of HPLC-DAD

| Parameter            | Condition                             |
|----------------------|---------------------------------------|
| Mobile phase         | Methanol : Acetonitrile (100:0 ; v/v) |
| Flow rate            | 1.00 mL/min                           |
| Column               | C18 (250 mm × 4.6 mm × 5 $\mu$ m )    |
| Detection wavelength | rutin 258, lutein 444 nm              |
| Retention time       | 20 min                                |
| $t_R$                | rutin = 2.31 min<br>lutein = 5.64 min |

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